



Dissolved organic carbon fluxes by seagrass meadows and macroalgal beds

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Estimates of dissolved organic carbon (DOC) release by marine macrophyte communities (seagrass meadows and macroalgal beds) based on *in situ* benthic chambers from published and unpublished are compiled in this study. The effect of temperature and light availability on DOC release by macrophyte communities was examined. Almost 85% of the seagrass communities and all of macroalgal communities examined acted as net sources of DOC. Net DOC fluxes in seagrass communities increase positively with water temperature. In macroalgal communities net DOC fluxes under light exceeded those under dark condition, however, this trend was weaker in seagrass communities. Shading of a mixed seagrass meadow in The Philippines led to a significant reduction on the net DOC release when shading was maintained for 6 days compared to only 2 days of shading. Net DOC fluxes increased with increasing community respiration, but were independent of primary production or net community production. The estimated global net DOC flux, and hence export, from marine macrophytes is about 0.158 ± 0.055 or 0.175 ± 0.056 Pg C year⁻¹ depending on the global extent of seagrass meadows considered.

Keywords: dissolved organic carbon (DOC), seagrass, macroalgae, coastal ecosystems, carbon export

INTRODUCTION

Dissolved organic carbon (DOC) represents only 2% of the carbon pool in the ocean, however, is the largest reservoir of exchangeable organic carbon in the marine carbon pool (685 Pg C; Hansell and Carlson, 2002). Marine primary producers release part of the carbon fixed photosynthetically as DOC, while the bulk of this organic carbon is consumed by bacteria (del Giorgio and Williams, 2005). Bacteria consume preferentially newly synthesized DOC, which is highly labile (Kenworthy and Thayer, 1984), however, a large portion of the bulk DOC pool in the marine ecosystem consists of refractory material which is transported in the ocean (Hansell et al., 2004). Thus, the production and consumption of DOC is a major step in the marine carbon cycle.

Marine macrophytes (i.e., macroalgae and seagrass) form highly productive ecosystems in the coastal ocean (Gattuso et al., 1998; Hemminga and Duarte, 2000; Barrón et al., 2006; Duarte et al., 2010). Marine macrophyte communities are complex assemblages including microbes, metazoans and plants, all of which are involved in dissolved organic matter (DOM) fluxes. These communities can release DOM by different mechanisms, such as release from autotrophic organisms (seagrass, benthic macro- and micro-algae, phytoplankton, and epiphytes), release by leaching, and decomposition of detritus and excretion by organisms all of which can be considered to be of autochthonous origin. DOM can also be produced from release

and/or decomposition of organic matter produced elsewhere (i.e., of allochthonous origin), such as DOM derived from decomposition of sestonic material trapped in the sediments (Gacia et al., 2002; Hendriks et al., 2008; Vaquer-Sunyer et al., 2012).

Marine macrophyte communities tend to produce excess organic matter that can be stored in sediments or exported to adjacent ecosystems (Duarte and Cebrián, 1996; Gattuso et al., 1998; Barrón et al., 2003; Duarte et al., 2005; Barrón and Duarte, 2009). Macroalgae and seagrasses store about 0.4 and 16% of their net primary production in the sediments, respectively (Duarte and Cebrián, 1996), and some of the excess organic matter they produce can be exported to adjacent waters (Duarte et al., 2005; Barrón and Duarte, 2009; Wada and Hama, 2013). The export of organic matter has been reported to account, on average, for 25 and 44% of the net primary production of macroalgae and seagrass, respectively (Duarte and Cebrián, 1996), although the form, particulate or dissolved, of this release has not been resolved yet. Release of dissolved organic carbon (DOC) by marine macrophytes has been examined mainly in experiments with isolated plants (Brilinsky, 1977; Wetzel and Penhale, 1979; Moriarty et al., 1986; Haas et al., 2010; Kaldy, 2012). DOC release rates of macrophytes isolated under laboratory conditions using ¹⁴C addition experiments, concluding that DOC release from macroalgae represented from 1 to 39% of gross primary production (Khailov and Burlakova, 1969; Brilinsky, 1977; Pregnall, 1983) compared to <5% from seagrasses (Brilinsky, 1977; Penhale and

Smith, 1977; Moriarty et al., 1986). These techniques, however, underestimate the release of DOC from the community because they measure only the carbon recently incorporated by the plant as DO^{14}C , while they neglect leaching from live and decaying tissues. Haas et al. (2010) reported a net DOC release of $1.3 \pm 0.5 \text{ mmol C m}^{-2} \text{ h}^{-1}$ and $0.2 \pm 0.2 \text{ mmol C m}^{-2} \text{ h}^{-1}$ using fragments of seagrass and algal specimens, respectively, incubated in beakers under natural daylight conditions. However, extrapolation of DOC release by individual plants to community level underestimates the magnitude of community net release, and hence total DOC export, because it neglects the release of DOC by other components of the community, such as epiphytes, the leaching of DOC from material decomposing in the sediments, and the consumption of DOC by bacteria and possibly metazoans. Indeed, some marine macrophyte communities may possibly act as DOC sinks, consuming more carbon than they produce, a possibility that cannot be ascertained from consideration of DOC release by individual plants alone. Thus, the capacity to export DOC is best assessed through the measurement of net DOC flux by the whole macrophyte community and, especially, using *in situ* incubations rather than laboratory experiments which do not damage the biological components of the community, ensuring reliable estimates of DOC flux.

Seasonal DOC release by marine macrophyte communities (Ziegler and Benner, 1999; Suzuki et al., 2003; Ziegler et al., 2004; Barrón and Duarte, 2009; Apostolaki et al., 2010a) suggests a dependence of DOC release on temperature and light availability. Metabolic rates were reported to be strongly temperature-dependent in a Mediterranean macroalgae meadow dominated by *Caulerpa prolifera*, as community respiration and gross primary production were higher when water temperature was around 28°C (Vaquer-Sunyer et al., 2012). Most marine macrophyte communities examined displayed net DOC release during light incubations and net uptake during dark (Maher and Eyre, 2010). Indeed, DOC release was on annual scale 12% higher in the light than in the dark in a *Posidonia oceanica* meadow (Apostolaki et al., 2010b). Furthermore, bacterial production in the water column of seagrass beds has been reported to vary diurnally (Chin-Leo and Benner, 1991). Additional evidence was provided by Ziegler and Benner (1999), who reported bacterial production and net benthic DOC production to vary between a sunny and a subsequent cloudy day. In addition, Maher and Eyre (2010) reported a tight coupling between DOC fluxes and bacterial biomass. Together, these results suggest a dependence of net DOC release on light availability.

Regrettably, studies examining the DOC flux by marine macrophyte communities *in situ* are still few (Ziegler and Benner, 1999; Barrón et al., 2003, 2004; Barrón and Duarte, 2009; Apostolaki et al., 2010a,b; Maher and Eyre, 2010). Here we experimentally quantify the net DOC flux by several marine macroalgal and seagrass communities around the world. We examine the effect of ambient water column temperature on net DOC fluxes of seagrass communities. The use of *in situ* benthic chambers could result in slight differences in the temperature in their interior. However, this minor inconvenient is superseded by the benefit of examining the whole community under near *in situ* conditions. We also test the effect of light availability on net community

DOC fluxes by comparing them under light and dark in short-term incubations and by examining the effect of shading on these fluxes in longer-term (days) incubations. We combine our results with our own published estimates as well as those of colleagues to provide an assessment of the global net DOC flux by marine macrophyte communities.

METHODS

NET DOC FLUXES FROM MARINE MACROPHYTES

The net DOC flux of seagrass and macroalgal communities was evaluated *in situ* using benthic chambers (3–4 replicates) set up by SCUBA divers. The benthic chambers consisted of a PVC cylinder (18 cm in diameter) inserted about 7–10 cm into the sediment and fitted with a polyethylene plastic bag by the divers (Hansen et al., 2000). The polyethylene plastic bags had a sampling port to collect the water samples with 50 ml acid-washed syringes. Most of the DOC fluxes during dark were collected from incubations spanning the entire night period, however, night incubations could not be set up in some locations. In these communities, dark DOC fluxes were estimated using parallel clear benthic chambers and benthic chambers covered with dark plastics bags. These incubations lasted around 2–3 h in The Philippines, around 4 h in Ria Formosa, and between 3 and 4 h in Florida. The volume of each chamber was estimated by injecting 5 ml concentrated phosphate solution (0.25 mol l^{-1}), and analyzing the water sample for phosphate concentration after mixing at the end of the incubation. The water samples collected were kept frozen until the spectrophotometric determination of the phosphate concentration (Hansen and Koroleff, 1999). The volume of the benthic chambers ranged from 5 to 20 l across sites. The volume of each benthic chamber was used to calculate the net DOC fluxes.

An aliquot of the water sample from the benthic chambers was fixed immediately for the determination of dissolved oxygen (DO), whereas the rest was filtered through pre-combusted (450°C for 4 h) GF/F filters into pre-combusted glass ampoules sealed under heat and analyzed later for DOC. DOC samples collected from March 2001 to May 2001 were kept frozen in acid washed material (glass vials encapsulated with silicone-teflon caps) until analyses. DOC samples collected subsequently were kept acidified with 2 N HCl at room temperature in acid-washed sealed ampoules. DOC samples were analyzed in a Shimadzu TOC-5000A (Benner and Strom, 1993). DOC standards provided by Dennis A. Hansell and Wenhao Chen (University of Miami) of 44–45 and $2 \mu\text{mol DOC}$ were used to assess the accuracy of the estimates. Samples for DOC analyses from the experiments conducted in The Philippines were kept frozen and analyzed by high temperature oxidation using an MQ1001 TOC Analyzer (Benner and Strom, 1993). Depending on the experimental location, DO concentration were determined by high-precision Winkler titration with a Mettler DL21 titrator (Carritt and Carpenter, 1966), with a reproducibility of 0.35%, and other samples were measured using a spectrophotometric modification of the Winkler titration method (Pai et al., 1993; Roland et al., 1999).

Fluxes of DOC were based on the rate of change in concentration between initial and final samples. Hourly dark and light DOC rates were converted to daily rates by multiplying the hourly rates by the number of night and light hours during the day.

Hourly rates of respiration (R) and net community production (NCP) were estimated from the difference in oxygen concentration change in the chambers during the night and the day, respectively. Gross primary production (GPP) was estimated as the sum of the hourly rates of R and NCP. Daily GPP rates were calculated using the photoperiod, daily R was calculated as hourly R multiplied by 24 h, and daily rates of NCP were estimated as the difference between daily GPP and R. Metabolic rates in DO units were converted to carbon units assuming photosynthetic and respiratory quotients of 1 (Kirk, 1983).

EFFECT OF TEMPERATURE ON NET DOC FLUXES

Ambient water column temperature was measured using a data logger (TidBit, Onset™) set up within the *P. oceanica* canopy in Magalluf Bay. Temperature was measured just above the canopy of *P. oceanica* in Sounion by means of a SEABIRD-19 CTD. When water temperature was not measured we searched for temperature values for the site sampled in the literature. For instance literature values were used for measurements in Bolinao, The Philippines (Gacia et al., 2005) and Delta del Ebro, Spain (Llebot et al., 2011).

EFFECT OF SHADING ON NET DOC FLUXES

The effect of shading on net community DOC release was examined in March 2000 in a mixed seagrass meadow at Silaqui Island, Bolinao (Luzón, The Philippines), and in October 2003 at a *Thalassia testudinum* meadow in Homossassa, on the Gulf coast of Florida.

Nine separate 2.25 m² (1.5 × 1.5 m) plots were set up in October 2003 in the *T. testudinum* meadow in Homossassa in the Gulf coast of Florida, experimental details are reported in Calleja et al. (2006). The shading screens (dark gray fiberglass) were established 60 cm above the seagrass canopy. Shade treatments were 0, 1, 21, 34, 48, 57, 84, and 86% light reduction on the eight plots. Benthic chambers were set up in the center of the plot to estimate the DOC fluxes of the *T. testudinum* community after 5 days of shading. The benthic chambers were deployed for 2.5–4 h during day and night. Net DOC fluxes were measured as described above.

Duplicate 2 m² plots with shading screens were set up in the mixed seagrass meadow at Silaqui Island (The Philippines), in each of the following treatments: 0, 50, 65, 84, 90, 95, and 98% of reduction of the light incident on the communities. Additional details on the experimental design are reported in Gacia et al. (2005). The DOC fluxes, after 2 and 6 days of shading, were estimated using transparent benthic chambers as described above. An additional series of four benthic chambers with thick black plastic bags fitted to the PVC ring were used to examine net DOC release in communities exposed to short-term (2–4 h) darkness.

GLOBAL DOC FLUXES

Estimates of net DOC fluxes in seagrass derived experimentally here were amended with estimates derived from the literature, resulting in a total of 73 estimates (Table 1), seven of which corresponded to DOC fluxes in macroalgal communities (Table 2). Weighted-mean daily net DOC fluxes by seagrass meadows and macroalgal communities per site were calculate when more than

one DOC flux estimate was reported (i.e., mean per DOC fluxes from *P. oceanica* in Mallorca and Greece). Standard errors of the weighted-average DOC fluxes were calculated using error propagation for each seagrass species per site. These weighted-mean net daily DOC fluxes per site were used to calculate the average net daily DOC fluxes, leading to 28 sites estimates from a total of 66 individual estimates in seagrass communities. A first order approximation of the global net DOC flux from marine macrophyte communities were derive as the product of the weighted average net daily DOC release by marine macrophytes and the global areal extent of seagrass meadows and macroalgal communities.

STATISTICAL ANALYSES

Least-squares linear regression analysis was used to describe the relationship between the net DOC fluxes and ambient water column temperature in seagrass communities, the net DOC fluxes under light and dark conditions in seagrass and macroalgae communities and between metabolism and net DOC fluxes in macrophyte communities. This analysis was also used to test the relationship between the light reduction and the net DOC fluxes in the shading experiments. We used the Wilcoxon signed rank test to test for tendencies for DOC fluxes in the light and dark to differ consistently. The Wilcoxon ranked sign test was also used to compare the net DOC fluxes after 2 and 6 days of shading in the experiment conducted in The Philippines.

RESULTS

The maximum net DOC uptake ($-53.6 \pm 10.8 \text{ mmol C m}^{-2} \text{ d}^{-1}$) and the maximum net release ($81.6 \pm 13.9 \text{ mmol C m}^{-2} \text{ d}^{-1}$) were both observed in *C. nodosa* communities (Table 1, Figure 1A). DOC release ranged an order of magnitude across macroalgal communities from $8.4 \pm 1.6 \text{ mmol C m}^{-2} \text{ d}^{-1}$ in *C. prolifera* to $71.9 \pm 33.2 \text{ mmol C m}^{-2} \text{ d}^{-1}$ in *Halimeda tuna* (Table 2, Figure 1B). The net DOC fluxes in seagrass communities were significantly correlated with the water temperature ($r^2 = 0.1$, $p < 0.05$), increasing by $1.5 \pm 0.7 \text{ mmol C m}^{-2} \text{ d}^{-1}$ by each °C temperature increase (Figure 2).

Light and dark net DOC flux were significantly correlated in seagrass ($r^2 = 0.17$, $p < 0.05$) and macroalgal ($r^2 = 0.72$, $p < 0.05$) communities (Figure 3). Net flux of DOC tended to be higher under light than dark conditions in macroalgal communities (Wilcoxon ranked sign test, $p < 0.05$; Figure 3A). However, although 30 out of 46 seagrass communities tended to support higher net DOC fluxes under light than under dark, this tendency was not statistically significant (Wilcoxon ranked sign test, $p > 0.05$; Figure 3A). An important fraction (40%) of the marine macrophyte communities studied had a net DOC flux under light conditions at least two fold higher than that in the dark (Figure 3B). There was no significant correlation between either GPP or NCP and net DOC fluxes for both seagrass and macroalgal communities ($p > 0.05$, Figures 4A,C). However, community R and net DOC flux were significantly correlated in seagrass communities ($r^2 = 0.12$, $p < 0.05$, Figure 4B), but not in macroalgal communities.

Experimentally-shaded *T. testudinum* communities in Homossassa (Florida) supported net DOC fluxes ranging from

Table 1 | Net dissolved organic carbon (DOC) fluxes (mean \pm SE; rates in mmol C m⁻² d⁻¹) in seagrass meadows.

Field site	Specie	Temp. °C	DOC \pm SE mmol C m ⁻² d ⁻¹	Comments	References
Bolinao, The Philippines	Mixed seagrass meadow	28.8	76.4	March, 2000	1, 9
	<i>Enhalus acoroides</i>	28.8	18.9 \pm 16.5	March, 2000	
Delta del Ebro, Spain	<i>Cymodocea nodosa</i>	26.5	-34.9 \pm 12.1	July, 2000, 0.5 years old	2, 10
		26.5	38.2 \pm 2.4	July, 2000, 3.3 years old	
		26.5	12.8 \pm 7.7	July, 2000, 2.3 years old	
		26.5	81.6 \pm 13.9	July, 2000, 6.1 years old	
		26.5	75.2 \pm 22.6	July, 2000, 3.8 years old	
		26.5	-53.6 \pm 10.8	July, 2000, 1.4 years old	
		26.5	17.0 \pm 8.9	July, 2000, 5.3 years old	
Mean			19.5 \pm 14.3		
Trondheim, Norway	<i>Zostera marina</i>		10.6 \pm 2.2	September, 2000	1, 11
			7.0 \pm 1.3	September, 2000	
Mean			8.8 \pm 1.4		
Porto Colom, Mallorca I., Spain	<i>P. oceanica</i>		1.8 \pm 1.4	May, 2001	1, 12
Cabrera I. Spain	<i>C. nodosa</i>		5.9 \pm 1.7	May, 2001	
Santa Maria, Cabrera I., Spain	<i>P. oceanica</i>		15.6 \pm 3.7	May, 2001	
Port, Cabrera I., Spain	<i>P. oceanica</i>		-1.0 \pm 2.3	May, 2001	
Cap Enderrocat, Mallorca I., Spain	<i>P. oceanica</i>		5.0 \pm 2.6	March, 2002	1
			13.8 \pm 3.9	June, 2002	
Mean			9.4 \pm 3.3		
Magaluf Bay, Mallorca I., Spain	<i>P. oceanica</i>		-18.5 \pm 12.2	June, 2002, at 22 m deep	1,13
			1.4 \pm 2.7	June, 2002, at 15 m deep	
			19.6 \pm 3.2	June, 2002, at 4 m deep	
Magaluf Bay, Mallorca I., Spain	<i>P. oceanica</i>		-20.8	March, 2001	3
			3.0 \pm 1.8	April	
			13.7 \pm 2.2	May	
		22.6	17.4 \pm 3.4	June	
		25.1	10.6	July	
		27.2	34.2 \pm 8.5	August	
		25.7	12.2 \pm 4.3	September	
		23.6	10.8 \pm 3.4	October	
		18.6	8.0 \pm 1.2	November	
		15.6	2.8 \pm 5.4	December	
		13.9	2.1 \pm 0.3	January, 2002	
		14.5	6.1 \pm 1.9	February	
		14.4	5.2 \pm 3.2	March	
		15.2	10.8 \pm 1.8	April	
		18.9	8.2 \pm 3.4	May	
		22.4	34.7 \pm 4.1	June	
		26.3	10.3 \pm 4.1	July	
		26.0	8.2 \pm 8.8	August	
		22.7	14.5 \pm 6.5	October	
Mean			11.4 \pm 3.3		
Ria formosa, Portugal	<i>Zostera noltii</i>		-13.3 \pm 9.7	July, 2002	4
	<i>C. nodosa</i>		-3.6 \pm 4.6	July, 2002	
Homossassa, Gulf of Florida, USA	<i>T. testudinum</i>		-0.2	October, 2003	1, 14
	<i>T. testudinum</i>		3.4	October, 2003	
Mean			1.6 \pm 1.3		

(Continued)

Table 1 | Continued

Field site	Specie	Temp. °C	DOC ± SE mmol C m ⁻² d ⁻¹	Comments	References
Homossassa, Gulf of Florida, USA	<i>T. testudinum</i>		-15.4 ± 3.5	October, 2003	1
	<i>Syringodium filiforme</i>		33.0 ± 10.0	October, 2003	
	<i>Halodule wrightii</i>		21.7 ± 32.1	October, 2003	
Laguna Madre, USA	<i>T. testudinum</i>	30.7	24.9 ± 7.9	July, 1996	5, 6
		30.5	23.0 ± 0.7	September	
		23.9	3.6 ± 19.6	November	
		11.6	3.5 ± 20.5	January, 1997	
		20.7	14.2 ± 11.7	March	
		27.3	5.6 ± 16.1	June	
Mean			12.5 ± 4.3		
Sounion Aegean Sea (Greece)	<i>P. oceanica</i>	13.9	4.6 ± 3.2	April, 2006	7
		23.4	5.0 ± 3.5	June	
		25.8	7.2 ± 2.6	August	
		21.3	12.6 ± 4.5	October	
		16.1	1.7 ± 1.1	December	
		14.0	-17.4 ± 2.7	February, 2007	
		15.5	6.4 ± 5.1	April	
Mean			2.9 ± 3.4		
Southeastern coast of Australia	<i>Posidonia</i> sp.		26.1 ± 3.7	Sandy sediments	8
	<i>Ruppia</i> sp.		22.4 ± 16.4	Muddy sediments	
	<i>Ruppia</i> sp.		26.1 ± 8.4	Muddy sediments	
	<i>Zostera</i> sp.		3.5 ± 9.0	Muddy sediments	
	<i>Zostera</i> sp.		41.5 ± 19.0	Sandy sediments	
	<i>Halophila</i> sp.		-8.1 ± 4.2	Muddy sediments	
	<i>Halophila</i> sp.		15.1 ± 11.4	Sandy sediments	
Average			12.4 ± 2.9		

Negatives values represent net uptake while positive values represent net release. Weighted mean DOC fluxes per site. Overall average of DOC fluxes are reported at the bottom. References are shown in the note.

Note. 1, This study; 2, Barrón et al., 2004; 3, Barrón and Duarte, 2009; 4, Santos et al., 2004; 5, Ziegler and Benner, 1999; 6, Ziegler et al., 2004; 7, Apostolaki et al., 2010a; 8, Maher and Eyre, 2010. Water column temperature 9, Gacia et al., 2005; 10, Llebot et al., 2011. Metabolism data: 11, Duarte et al., 2002; 12, Holmer et al., 2004; 13, Gazeau et al., 2005; 14, Calleja et al., 2006.

a net uptake of $-10.4 \text{ mmol C m}^{-2} \text{ d}^{-1}$ to a net DOC release of $31.9 \text{ mmol C m}^{-2} \text{ d}^{-1}$ (Table 3), independently of the level of shading imposed ($r^2 = 0.13$, $p > 0.05$). The mixed seagrass meadow in The Philippines acted as a source of DOC under all irradiances after 2 days of shading, except for a small net DOC uptake of $-25.3 \text{ mmol C m}^{-2} \text{ d}^{-1}$ at 88.2% of light reduction. However, after 6 days of shading all the communities receiving $>10\%$ of incident light (7 plots in total) reverted to net DOC uptake, and only a net DOC release of $284.9 \text{ mmol C m}^{-2} \text{ d}^{-1}$ persisted at 98% shading (Figure 5). Net DOC flux in the experimentally-shaded mixed meadow was independent of the level of shading imposed after 2 days of shading ($r^2 = 0.11$, $p > 0.05$) and after 6 days of shading ($r^2 = 0.11$, $p > 0.05$). However, the net DOC fluxes in this seagrass meadow were significantly reduced after 6 days of experimental shading (Wilcoxon ranked sign test, $p < 0.05$) compared to net DOC fluxes after 2 days of shading.

The net DOC flux by seagrass meadows averaged $12.4 \pm 2.9 \text{ mmol C m}^{-2} \text{ d}^{-1}$ (Table 1), with 83% of the seagrass meadows tested acting as sources of DOC to the surrounding water (Figure 1A). All macroalgal communities studied acted as net sources of DOC (Table 2, Figure 1B), with a higher average ($\pm SE$) net DOC flux of $23.2 \pm 12.6 \text{ mmol C m}^{-2} \text{ d}^{-1}$ than that supported by seagrass meadows.

DISCUSSION

Published measurements of DOC release by marine macrophyte communities were few prior to the present study (Tables 1, 2). Maher and Eyre (2010) reported net DOC release by several seagrass communities studied in the South-eastern coast of Australia ranging from 3.5 to $41.5 \text{ mmol C m}^{-2} \text{ d}^{-1}$, except for a net DOC uptake of $8.1 \text{ mmol C m}^{-2} \text{ d}^{-1}$ in *Halophila* sp. (Table 1). Ziegler and Benner (1999) reported a consistent net DOC release by a *T. testudinum* community in the Gulf of Mexico, ranging

Table 2 | Net dissolved organic carbon (DOC) fluxes (mean \pm SE; rates in mmol C m⁻² d⁻¹) in macroalgal communities.

Field site	Specie	DOC \pm SE mmol C m ⁻² d ⁻¹	Comments	References
Bolinao, The Philippines	<i>Caulerpa racemosa</i>	37.7 \pm 6.2	March, 2000	1
	<i>Halimeda tuna</i>	71.9 \pm 33.1	March, 2000	
Oslofjord, Norway	<i>F. serratus</i>	35.2 \pm 5.5	August, 2000	1
Trondheim, Norway	<i>Laminaria</i>	30.6	September, 2000	1
	<i>Fucus vesiculosus</i>	15.0 \pm 1.9	September, 2000	
Porto Colom, Mallorca I., Spain	<i>C. prolifera</i>	8.4 \pm 1.6	May, 2001	1, 3
Southeastern coast of Australia	Habitat dominated by macroalgae	10.1 \pm 5.2		2
Average		23.2 \pm 12.6		

Positive values represent a net release. References are shown in the note.

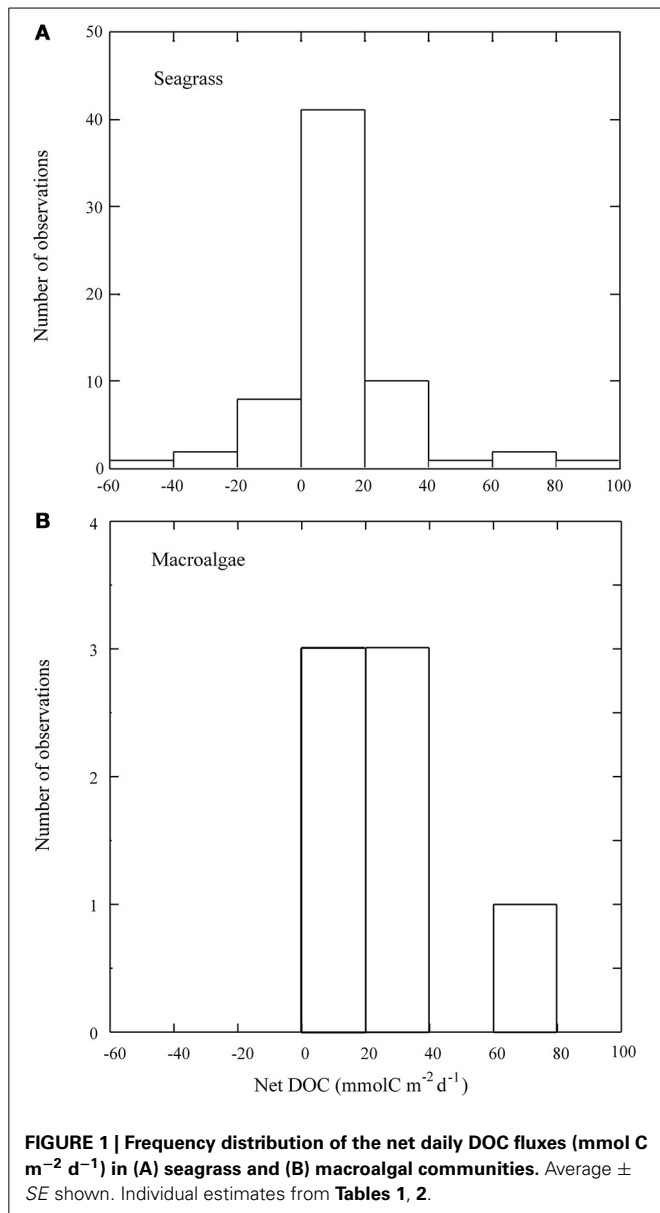
Note: 1, This study; 2, Maher and Eyre, 2010. Metabolism data; 3, Holmer et al., 2004.

from 3.6 to 24.9 mmol C m⁻² d⁻¹, which were substantially higher than the estimates of net DOC fluxes in the *T. testudinum* community studied here (Table 1). Velimirov (1986) reported a net DOC release by a Mediterranean *P. oceanica* community of 40.2 mmol C m⁻² d⁻¹ (Velimirov, 1986), which must be considered with caution as DOC concentrations in this report varied by almost 10 fold. Most (90%) of the experiments conducted in *P. oceanica* communities supported a net DOC release, with a maximum DOC release of 34.7 mmol C m⁻² d⁻¹. Barrón et al. (2004) reported a transition of communities of the seagrass *C. nodosa* in a Mediterranean Bay from a net sink to net source of DOC along the colonization process. The results presented show that marine macrophyte communities generally act as net sources of DOC to the ecosystem, 85% of the relative communities studied supported a net DOC release. Rates for individual net DOC fluxes in seagrass communities vary greatly between a net DOC uptake of 53.6 \pm 10.8 mmol C m⁻² d⁻¹ to a net DOC release of 81.6 \pm 13.9 mmol C m⁻² d⁻¹. Multiple components of the marine macrophyte communities (including microbes, metazoans, and plants) might be responsible for a net DOC uptake in seagrass communities. For instance, planktonic respiration might be enhanced within the canopy and could lead to a net DOC uptake in seagrass communities. In addition, metazoans can also consume DOC, a possibility well established for sponges, for which DOC uptake may provide around 80% of their organic carbon demand (Mueller et al., 2014). Furthermore, Vonk et al. (2008) using stable isotopes report that seagrass leaves take up dissolved organic nitrogen, thereby implying the uptake of DOC, but this has uptake by seagrass leaves has never been investigated directly. Likewise, net DOC release maybe supported by multiple components, including the plants themselves, epiphytic community, benthic micro- algae, phytoplankton, plant litter, metazoans, and release from sediment organic carbon. However, the relative contribution of each component to the net DOC uptake has not yet been apportion for seagrass communities, which is, therefore, an open question that requires further attention. Published reports of DOC release by macroalgal communities were few. Net DOC release by a macroalgal-dominated community in the

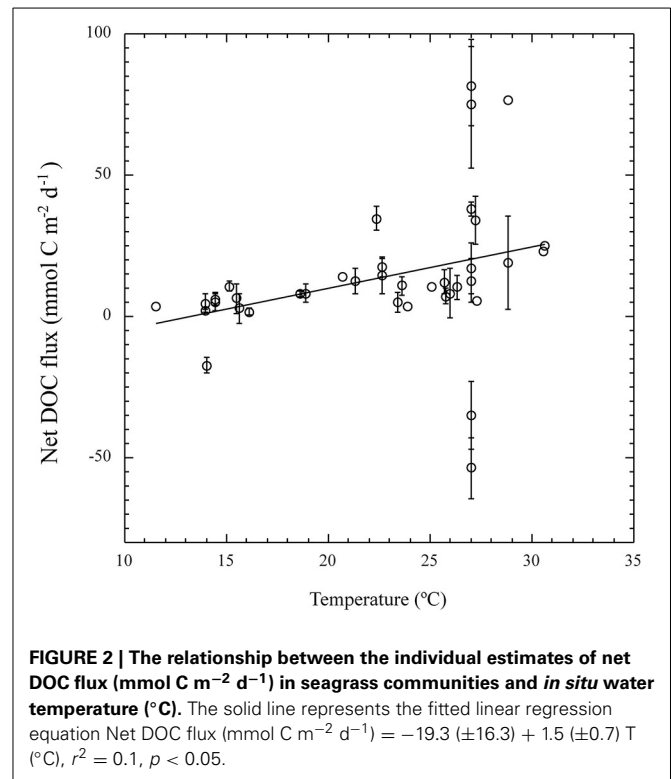
South-eastern coast of Australia was 10.1 \pm 5.2 mmol C m⁻² d⁻¹ (Maher and Eyre, 2010). In our study, all the macroalgal communities were net sources of DOC to the ecosystem ranging from 8.4 \pm 1.6 to 71.9 \pm 33.1 mmol C m⁻² d⁻¹. Wada and Hama (2013) report that macroalgae contribute up to 20% of total DOC concentration in Oura Bay in Japan.

Ziegler and Benner (1999) showed that the net DOC fluxes in a *T. testudinum* community were higher in the light, ranging between a release of 8.4–22.9 mmol C m⁻² d⁻¹, while in the dark DOC fluxes varied from a net uptake of 7.7 to a net release of 8.4 mmol C m⁻² d⁻¹, and that DOC fluxes under the light being significantly correlated with NCP. Our results also revealed a tendency for marine macrophyte communities to support higher net DOC fluxes under the light than in the dark, a tendency that was particularly strong in macroalgae communities. *P. oceanica* communities had higher net DOC release rates in summer than in winter when water temperature was higher (Barrón and Duarte, 2009), despite the increase in bacterial carbon demand with increased temperature (Kritzberg et al., 2010). The absence of any significant relationship between net DOC fluxes and GPP or NCP across communities suggests a weak coupling between light availability and community photosynthetic activity or net community DOC release. However, there was a significant relationship between community respiration and net DOC fluxes, as reported previously for a *C. nodosa* community (Barrón et al., 2004). Several studies link community respiration and DOC fluxes in marine macrophyte communities. Community respiration in a Mediterranean macroalgal community dominated by *C. prolifera* is driven by the release of DOC (Vaquer-Sunyer et al., 2012). Maie et al. (2006) reported high DOC release during decomposition of the seagrass *T. testudinum*, with 80% of the DOC leached over the first week of decomposition. This suggests that the relationship between community respiration rate and net DOC release rate may derive from high DOC release and respiration rates.

Further insight into the relationship between incident irradiance, photosynthetic activity, and net DOC release was derived from the two shading experiments conducted here. Net DOC

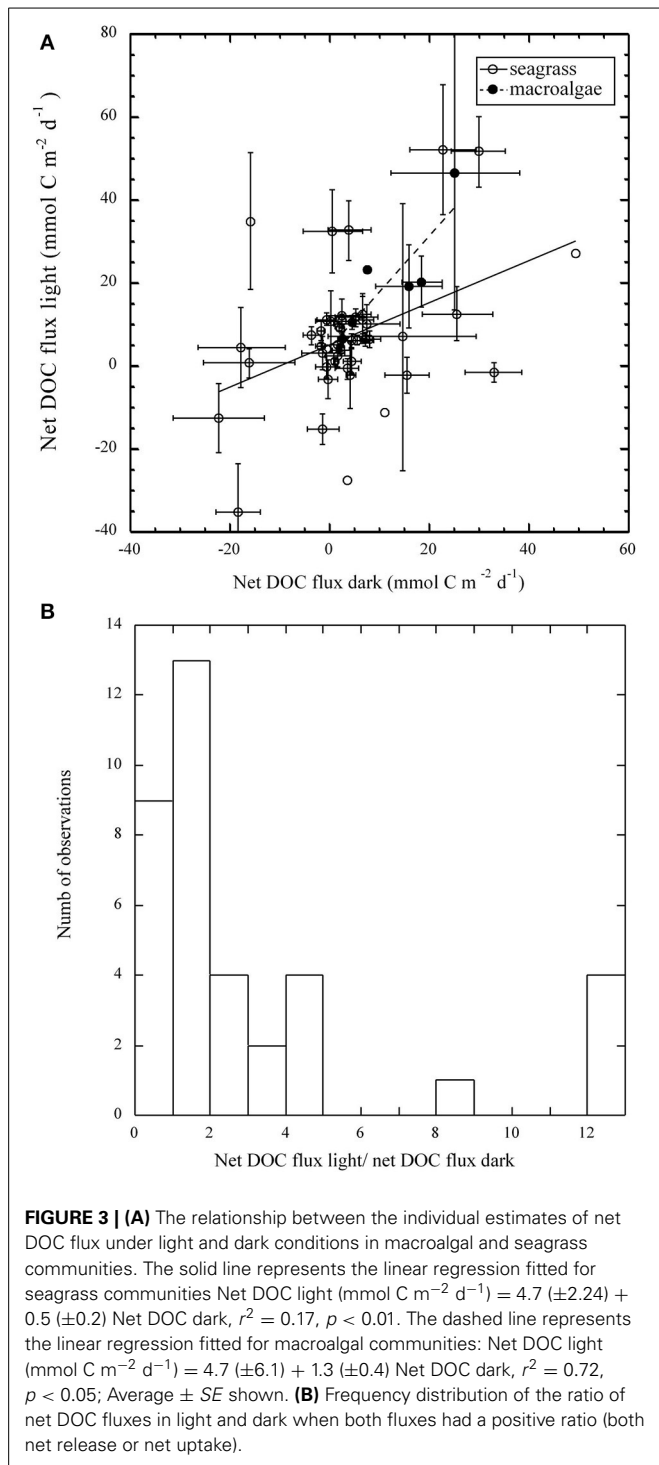


fluxes were not affected by shading for 5 days of a *T. testudinum* community, whereas the GPP and NCP, but not respiration rates of this seagrass meadow were significantly reduced (Calleja et al., 2006). Shading of a mixed seagrass meadow in The Philippines did not lead to any significant reduction in net DOC release after 2 days of shading, but it led to a significant reduction of the net DOC release when shading was maintained for 6 days. These results suggest, therefore, that whereas net community DOC release is not supported by fresh photosynthetic activity, the necessary coupling between photosynthetic production and DOC release occurs at time scales of a few days in seagrass meadows. The reduction in net DOC fluxes after 6 days of shading could have been caused by a reduction of the net DOC release from seagrass communities or by an increase in DOC consumption due to mainly bacterial respiration as suggested above.



DOC release by a *T. testudinum* community represented about 10% of the net community production in Laguna Madre (Ziegler and Benner, 1999). The rates of net DOC release by seagrass communities reported here represented on average 38% of the GPP, but 83% of their NCP. The net DOC release in macroalgal communities represented on average 27% of the GPP but more than 300% of the NCP. Because carbon burial and particulate carbon export by macrophyte beds typically exceed 40% of NPP (Duarte and Cebrián, 1996), the high net DOC release relative to NCP of the macrophyte communities studied here is unlikely to be supported by autochthonous photosynthetic production alone (Barrón et al., 2004). Recently, Maher and Eyre (2011) reported that the $\delta^{13}\text{C}$ -DOC value from estuarine water column is similar to the seagrass and algal ^{13}C values indicating that the DOC pool was dominated by macrophyte-derived DOC. However, inputs and subsequent decay of seston in seagrass sediments (e.g., Gacia et al., 2002; Hendriks et al., 2008; Kennedy et al., 2010), may play an important role in supporting the net DOC release of macrophyte communities. Use of stable isotopes to explore the link between macrophyte communities and DOC pools will help to further resolve the sources of DOC fluxes, autochthonous or allochthonous, in these communities.

The total global net DOC flux from marine macrophytes was estimated using the weighted average net DOC release by marine macrophytes derived here of 4521.6 ± 1073.1 mmol C m⁻² year⁻¹ and 8457.1 ± 4602.7 mmol C m⁻² year⁻¹ for seagrass meadows and macroalgal communities, respectively, and the area estimated to be covered by macroalgae (1.4×10^{12} m², Duarte et al., 2005) and seagrass meadows [0.3×10^{12} m²].



(Duarte et al., 2005) to $0.6 \times 10^{12} \text{ m}^2$ (Charpy-Roubaud and Sournia, 1990)]. The estimated global net DOC flux, and hence export, from seagrass meadows ranged from $0.016 \pm 0.004 \text{ Pg C year}^{-1}$ to $0.033 \pm 0.008 \text{ Pg C year}^{-1}$ depending on whether the low or high estimates of global seagrass cover is used, respectively. This range encompasses the global net DOC flux of $0.019 \text{ Pg C year}^{-1}$ estimated by Maher and Eyre (2010). The global net DOC export from seagrass meadows reported here represents 46% of

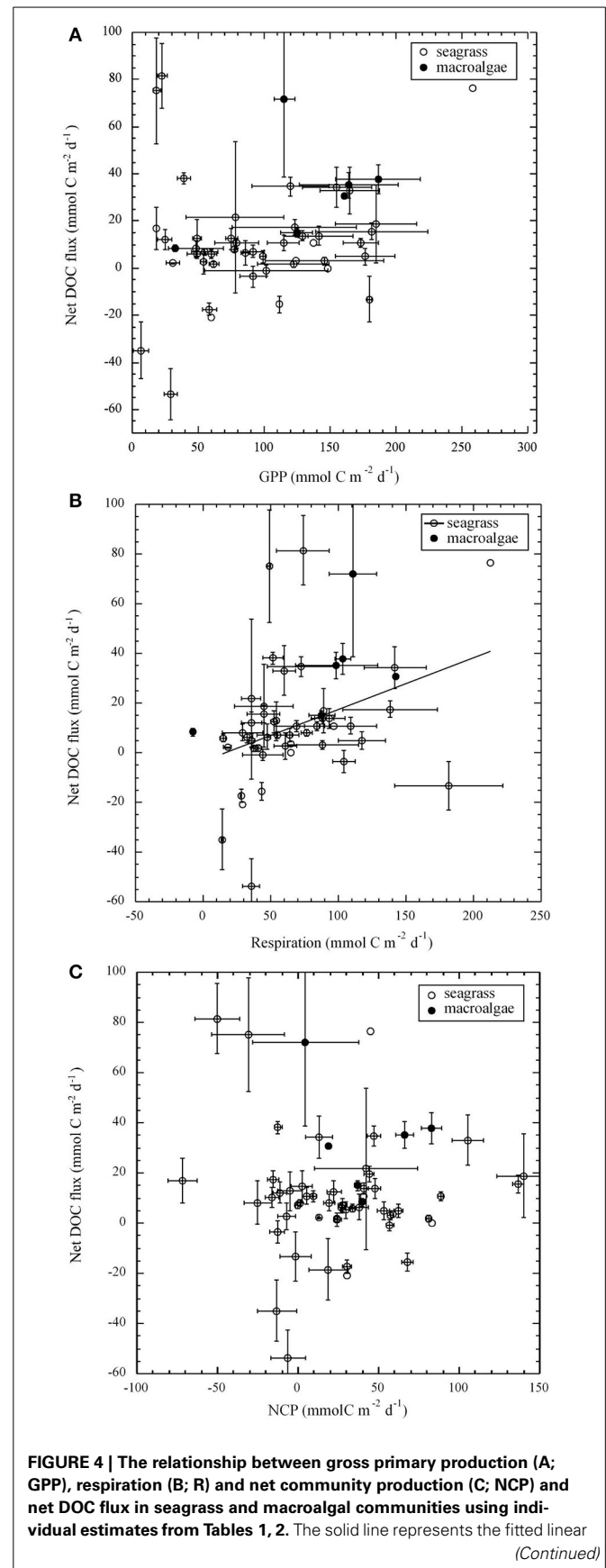


FIGURE 4 | Continued

regression equation between respiration and net DOC flux in seagrasses;
 Net DOC flux ($\text{mmol C m}^{-2} \text{ d}^{-1}$) = $-2.5 (\pm 6.3) + 0.2 (\pm 0.1) R$, $r^2 = 0.12$,
 $p < 0.05$.

Table 3 | Net DOC flux ($\text{mmol C m}^{-2} \text{ d}^{-1}$) in the *Thalassia testudinum* meadow after 5 days of shading (% light reduction) in Florida.

% Light reduction	Net DOC fluxes $\text{mmol C m}^{-2} \text{ d}^{-1}$
0	3.4
1	-0.2
21	-10.4
34	31.9
48	4.3
57	-5.0
84	20.4
86	11.0

Negative values represent net DOC uptake.

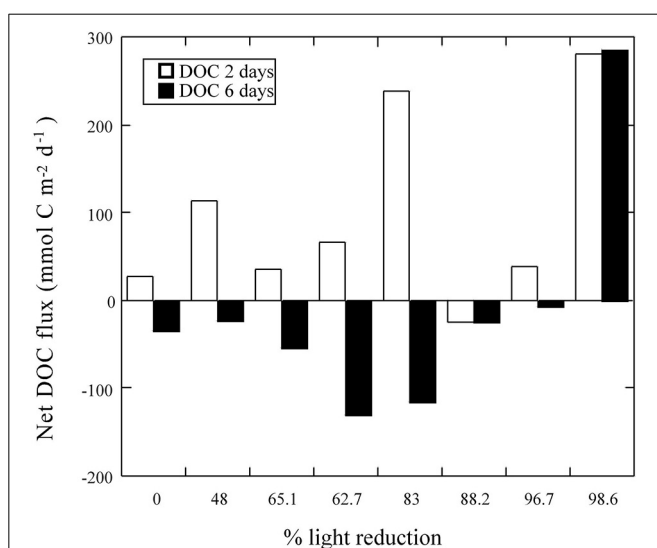


FIGURE 5 | The relationship between the net DOC flux under light conditions ($\text{mmol C m}^{-2} \text{ d}^{-1}$) and the experimental shading in the mixed Philippine seagrass meadow after 2 days of shading; Net DOC fluxes ($\text{mmol C m}^{-2} \text{ d}^{-1}$) = $20.6 (\pm 94.96) + 1.1 (\pm 1.3)\%$ light reduction, $r^2 = 0.11$, $p > 0.05$, and 6 days of shading Net DOC fluxes ($\text{mmol C m}^{-2} \text{ d}^{-1}$) = $-102.8 (\pm 112.3) + 1.3 (\pm 1.5)\%$ light reduction, $r^2 = 0.11$, $p > 0.05$. Negative values represent net DOC consumption.

the global NCP of seagrass meadows calculated by Duarte et al. (2010). This high export of dissolved organic carbon in relation to the seagrass NCP could be partly supported by allochthonous carbon inputs, derived from the capacity of seagrass canopies to trap particulate material from the water column (e.g., Hendriks et al., 2008). Indeed, in general, half of the organic carbon in seagrass sediments is of allochthonous origin (Kennedy et al., 2010), some of which could be released as DOC. The estimated

global net DOC export from macroalgal beds is $0.14 \pm 0.08 \text{ Pg C year}^{-1}$, well above that of seagrass meadows and in the range of other global DOC export estimates from macroalgae beds estimated at $0.061 \text{ Pg C year}^{-1}$ (Maher and Eyre, 2010) and $0.34 \text{ Pg C year}^{-1}$ (Hughes et al., 2012). Thus, the global net DOC release by marine macrophytes supports a net carbon export of about $0.158 \pm 0.055 \text{ Pg C year}^{-1}$ to $0.175 \pm 0.056 \text{ Pg C year}^{-1}$. We acknowledge that this estimate bears considerable uncertainties. First, the total number of net DOC flux estimates from marine macrophytes available is still limited, particularly in the Southern Hemisphere. For instance, only 11% of the individual estimates of net DOC fluxes from seagrass communities are from the Southern Hemisphere, specifically from Southeastern coast of Australia. In addition, a second source of uncertainty is that this study represents only the net DOC fluxes from 10 seagrass species addressing that more than 40 of seagrass species have not been studied yet in relation to its net DOC fluxes. This lack of knowledge is even higher as we report just 7 estimates of net DOC fluxes out of the thousands macroalgae species. We acknowledge that the available data are still limited in terms of the total number of communities investigated, their geographic spread and their suitability to represent an annual flux. Hence, further efforts are needed to extend and increase the estimates of net DOC fluxes from marine macrophytes around the world, particularly in the Southern Hemisphere where net DOC fluxes from marine macrophytes are particularly scarce.

Despite the uncertainties described above, the global estimate of the net DOC release by marine macrophytes derived here provides a first-order approximation sufficient to assess whether macrophyte-derived DOC maybe a globally significant C flux in the ocean, thereby helping raise awareness on the importance of this process. The global net DOC export from these particular benthic ecosystems represents a small amount of the DOC pool in the ocean, however, it represents more than 50% of the annual global river input of DOC to the ocean that has been estimated to be 0.25 Pg C (Cauwet, 2002). The global net DOC export from macrophyte communities represents a modest flux relative to the metabolism of the coastal ocean, but is significant relative to the net community production of benthic habitats and of a similar magnitude as organic carbon burial rates in coastal sediments (Duarte et al., 2005). The fate of the DOM exported from marine macrophyte communities is, however, unknown. DOM release represents an important, but poorly understood flux from macrophyte communities that may contribute to support metabolism in adjacent pelagic ecosystems. This potential carbon input to the open ocean might be altered due to anthropogenic pressures in the coastal ecosystem with different outcomes in the benthic ecosystems. Globally, seagrass area has declined by over 29% (Waycott et al., 2009), however, macroalgae abundance has increase in coral reefs (Halpern et al., 2008) and some eutrophied environments. Consequently, net DOC fluxes from marine macrophytes in the coastal zone might increase in the near future due to the proliferation of macroalgae overcompensating for the decline in seagrass meadows if current human pressures in the coastal zone persists.

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REFERENCES

- Apostolaki, E. T., Holmer, M., Marba, N., and Karakassis, I. (2010a). Metabolic imbalance in coastal vegetated (*Posidonia oceanica*) and unvegetated benthic ecosystems. *Ecosystems* 13, 459–471. doi: 10.1007/s10021-010-9330-9
- Apostolaki, E. T., Holmer, M., Marba, N., and Karakassis, I. (2010b). Degrading seagrass (*Posidonia oceanica*) ecosystems: a source of dissolved matter to the Mediterranean. *Hydrobiologia* 649, 13–23. doi: 10.1007/s10750-010-0255-2
- Barrón, C., and Duarte, C. M. (2009). Dissolved organic matter release in *Posidonia oceanica* meadow. *Mar. Ecol. Prog. Ser.* 374, 75–84. doi: 10.3354/meps07715
- Barrón, C., Duarte, C. M., Frankignoulle, M., and Borges, A. V. (2006). Organic carbon metabolism and carbonate dynamics in a Mediterranean seagrass (*Posidonia oceanica*) meadow. *Est. Coasts* 29, 417–426. doi: 10.1007/BF02784990
- Barrón, C., Marbà, N., Duarte, C. M., Pedersen, M. F., Lindblad, C., Kersting, K., et al. (2003). High organic carbon export precludes eutrophication effects in experimental rocky shore communities. *Ecosystems* 6, 144–153. doi: 10.1007/s10021-002-0402-3
- Barrón, C., Marbà, N., Terrados, J., Kennedy, H., and Duarte, C. M. (2004). Community metabolism and carbon budget along a gradient of seagrass (*Cymodocea nodosa*) colonization. *Limnol. Oceanogr.* 49, 1642–1651. doi: 10.4319/lo.2004.49.5.1642
- Benner, R., and Strom, M. (1993). A critical evaluation of the analytical blank associated with DOC measurements by high-temperature catalytic oxidation. *Mar. Chem.* 41, 153–160. doi: 10.1016/0304-4203(93)90113-3
- Brilinsky, M. (1977). Release of dissolved organic matter by some marine macrophytes. *Mar. Biol.* 39, 213–220. doi: 10.1007/BF00390995
- Calleja, M., Ll., Barrón, C., Hale, J., Notestein, S., Vaughan, D., Frazer, T., et al. (2006). Light regulation of benthic sulfate reduction rates mediated by seagrass (*Thalassia testudinum*) metabolism. *Est. Coasts* 29, 1255–1264. doi: 10.1007/BF02781825
- Carritt, D. E., and Carpenter, J. H. (1966). Comparison and evaluation of currently employed modifications of the Winkler method for determining dissolved oxygen in seawater; a NASCO Report. *J. Mar. Res.* 24, 286–319.
- Cauwet, G. (2002). "DOM in the coastal zone," in *Biogeochemistry of Marine Dissolved Organic Matter*, eds D. A. Hansell and C. A. Carlson (San Diego, CA: Academic Press), 579–609.
- Charpy-Roubaud, C., and Sournia, A. (1990). The comparative estimation of phytoplanktonic and microphytobenthic production in the oceans. *Mar. Microb. Food Webs* 4, 31–57.
- Chin-Leo, G., and Benner, R. (1991). Dynamics of bacterioplankton abundance and production in seagrass communities of a hypersaline lagoon. *Mar. Ecol. Prog. Ser.* 73, 219–230. doi: 10.3354/meps073219
- del Giorgio, P. A., and Williams, P. J. B. (2005). "The global significance of respiration in aquatic ecosystems: from single cells to the biosphere," in *Respiration in Aquatic Ecosystems, Chapter 14*. (Oxford: Oxford University Press). doi: 10.1093/acprof:oso/9780198527084.003.0014
- Duarte, C. M., and Cebrián, J. (1996). The fate of marine autotrophic production. *Limnol. Oceanogr.* 41, 1758–1766. doi: 10.4319/lo.1996.41.8.1758
- Duarte, C. M., Marbà, N., Gacia, E., Fourqurean, J. W., Beggins, J., Barrón, C., et al. (2010). Seagrass community metabolism: assessing the carbon sink capacity of seagrass meadows. *Glob. Biogeochem. Cycle* 24, GB4032. doi: 10.1029/2010GB003793
- Duarte, C. M., Martínez, R., and Barrón, C. (2002). Biomass, production and rhizome growth near the northern limit of seagrass (*Zostera marina*) distribution. *Aquat. Bot.* 72, 183–189. doi: 10.1016/S0304-3770(01)00225-X
- Duarte, C. M., Middelburg, J. J., and Caraco, N. (2005). Major role of marine vegetation on the oceanic carbon cycle. *Biogeosciences* 2, 1–8. doi: 10.5194/bg-2-1-2005
- Gacia, E., Duarte, C. M., and Middelburg, J. J. (2002). Carbon and nutrient deposition in a Mediterranean seagrass (*Posidonia oceanica*) meadow. *Limnol. Oceanogr.* 47, 23–32. doi: 10.4319/lo.2002.47.1.0023
- Gacia, E., Kennedy, H., Duarte, C. M., Terrados, J., Marbà, N., Papadimitriou, S., et al. (2005). Light-dependence of the metabolic balance of a highly productive Philippine seagrass community. *J. Exp. Mar. Biol. Ecol.* 316, 55–67. doi: 10.1016/j.jembe.2004.10.008
- Gattuso, J.-P., Frankignoulle, M., and Wollast, R. (1998). Carbon and carbonate metabolism in coastal aquatic ecosystems. *Annu. Rev. Ecol. Syst.* 29, 405–434. doi: 10.1146/annurev.ecolsys.29.1.405
- Gazeau, F., Duarte, C. M., Gattuso, J.-P., Barrón, C., Navarro, N., Ruiz, S., et al. (2005). Whole-system metabolism and CO₂ fluxes in a Mediterranean Bay dominated by seagrass beds (Palma Bay, NW Mediterranean). *Biogeosciences* 2, 43–60. doi: 10.5194/bg-2-43-2005
- Haas, A. F., Jantzen, C., Naumann, M. S., Iglesias-Prieto, R., and Wild, C. (2010). Organic matter release by the dominant primary producers in a Caribbean reef lagoon: implication for *in situ* O₂ availability. *Mar. Ecol. Prog. Ser.* 409, 27–39. doi: 10.3354/meps08631
- Halpern, B. S., Walbridge, S., Selkoe, K. A., Kappel, C. V., Micheli, F., D'Agrosa, C., et al. (2008). A global map of human impact on marine ecosystems. *Science* 319, 948–952. doi: 10.1126/science.1149345
- Hansell, D. A., and Carlson, C. A. (2002). *Biogeochemistry of Marine Dissolved Organic Matter*. San Diego, CA: Academic Press.
- Hansell, D. A., Kadko, D., and Bates, N. R. (2004). Degradation of terrigenous dissolved organic carbon in the western Arctic ocean. *Science* 304, 858–861. doi: 10.1126/science.1096175
- Hansen, H. P., and Koroleff, F. (1999). "Determination of nutrients," in *Methods of Seawater Analysis*, eds K. Grasshoff, K. Kremling, and M. Ehrhardt (Weinheim: Wiley-VCH), 170–193.
- Hansen, J. W., Thamdrup, B., and Jørgensen, B. B. (2000). Anoxic incubation of sediment in gas-tight plastic bags: a method for biogeochemical process studies. *Mar. Ecol. Prog. Ser.* 208, 273–282. doi: 10.3354/meps208273
- Hemminga, M., and Duarte, C. M. (2000). *Seagrass Ecology*. Cambridge: Cambridge University Press.
- Hendriks, I. E., Sintès, T., Bouma, T., and Duarte, C. M. (2008). Experimental assessment and modeling evaluation of the effects of seagrass (*P. oceanica*) on flow and particle trapping. *Mar. Ecol. Prog. Ser.* 356, 163–173. doi: 10.3354/meps07316
- Holmer, M., Duarte, C. M., Boschker, H. T. S., and Barrón, C. (2004). Carbon cycling and bacterial carbon sources in pristine and impacted Mediterranean seagrass sediments. *Aquat. Microb. Ecol.* 36, 227–237. doi: 10.3354/ame036227
- Hughes, A. D., Black, K. D., Campbell, I., Davidson, K., Kelly, M. S., and Stanley, M. S. (2012). Does seaweed offer a solution for bioenergy with biological carbon capture and storage? *Greenhouse Gas Sci. Technol.* 2, 402–407. doi: 10.1002/ghg
- Kaldy, J. (2012). Influence of light, temperature and salinity on dissolved organic carbon exudation rates in *Zostera marina* L. *Aquat. Biosyst.* 8:19. doi: 10.1186/2046-9063-8-19
- Kennedy, H., Beggins, J., Duarte, C. M., Fourqurean, J. W., Holmer, M., Marbà, N., et al. (2010). Seagrass sediments as a global carbon sink: isotopic constraints. *Global Biogeochem. Cycles* 24, GB4026. doi: 10.1029/2010GB003848
- Kenworthy, W. J., and Thayer, G. W. (1984). Production and decomposition of the roots and rhizomes of seagrasses, *Zostera marina* and *Thalassia testudinum*, in temperate and subtropical marina ecosystems. *Bull. Mar. Sci.* 35, 364–379.
- Khailov, K. M., and Burlakova, Z. P. (1969). Release of dissolved organic matter by marine seaweeds and distribution of their total organic production to inshore communities. *Limnol. Oceanogr.* 14, 521–532.

- Kirk, J. T. O. (1983). *Light and Photosynthesis in Aquatic Ecosystems*. Cambridge: Cambridge University Press.
- Kritzbeg, E. S., Arrieta, J. M., and Duarte, C. M. (2010). Temperature and phosphorus regulating carbon flux through bacteria in a coastal marine system. *Aquat. Microb. Ecol.* 58, 141–151. doi: 10.3354/ame01368
- Llebot, C., Solé, J., Delgado, M., Fernández-Tejedor, M., Camp, J., and Estrada, M. (2011). Hydrographical forcing and phytoplankton variability in two semi-enclosed estuarine bays. *J. Mar. Syst.* 86, 69–86. doi: 10.1016/j.jmarsys.2011.01.004
- Maher, D. T., and Eyre, B. D. (2010). Benthic fluxes of dissolved organic carbon in three temperate Australian estuaries: implications for global estimates of benthic DOC fluxes. *J. Geophys. Res.* 115, G04039. doi: 10.1029/2010JG001433
- Maher, D. T., and Eyre, B. D. (2011). Insights into estuarine benthic dissolved organic carbon (DOC) dynamics using $\delta^{13}\text{C}$ -DOC values, phospholipid fatty acids and dissolved organic nutrient fluxes. *Geochim. Cosmochim. Acta* 75, 1889–1902. doi: 10.1016/j.gca.2011.01.007
- Maie, N., Jaffé, R., Miyoshi, T., and Childers, D. L. (2006). Quantitative and qualitative aspects of dissolved organic carbon leached from senescent plants in an oligotrophic wetland. *Biogeochemistry* 78, 285–314. doi: 10.1007/s10533-005-4329-6
- Moriarty, D. J. W., Iverson, R. L., and Pollard, P. C. (1986). Exudation of organic carbon by the seagrass *Halodule wrightii* Aschers and its effect on bacterial growth in the sediment. *J. Exp. Mar. Biol. Ecol.* 96, 115–126. doi: 10.1016/0022-0981(86)90237-6
- Mueller, B., van der Zande, R. M., van Leent, P. J. M., Meesters, E. H., Vermeij, M. J. A., and van Duyl, F. C. (2014). Effect of light availability on dissolved organic carbon release by Caribbean reef algae and corals. *Bull. Mar. Sci.* 90, 875–893. doi: 10.5343/bms.2013.1062
- Pai, S.-C., Gong, G.-C., and Liu, K.-K. (1993). Determination of dissolved oxygen in seawater by direct spectrophotometry of total iodine. *Mar. Chem.* 41, 343–351. doi: 10.1016/0304-4203(93)90266-Q
- Penhale, P. A., and Smith, W. O. (1977). Excretion of dissolved organic carbon by eelgrass (*Zostera marina*) and its epiphytes. *Limnol. Oceanogr.* 22, 400–407. doi: 10.4319/lo.1977.22.3.0400
- Pregall, A. M. (1983). Release of dissolved organic carbon from the estuarine intertidal macroalga *Enteromorpha prolifera*. *Mar. Biol.* 73, 37–42. doi: 10.1007/BF00396283
- Roland, F., Caraco, N. F., Cole, J. J., and del Giorgio, P. (1999). Rapid and precise determination of dissolved oxygen by spectrophotometry: evaluation of interference from color and turbidity. *Limnol. Oceanogr.* 44, 1148–1154. doi: 10.4319/lo.1999.44.4.1148
- Santos, R., Silva, J., Alexandre, A., Navarro, N., Barrón, C., and Duarte, C. M. (2004). Ecosystem metabolism and carbon fluxes of a tidal-dominated coastal lagoon. *Estuaries* 27, 977–985. doi: 10.1007/BF02803424
- Suzuki, Y., Fujii, M., Casareto, B. E., Furuta, A., and Ishikawa, Y. (2003). CO_2 sequestration and fate of organic matter with seagrass (*Zostera marina*) ecosystems. *J. Chem. Eng. Jpn.* 36, 417–427. doi: 10.1252/jcej.36.417
- Vaquier-Sunyer, R., Duarte, C. M., Jorda, G., and Ruiz-Halpern, S. (2012). Temperature dependence of oxygen dynamics and community Metabolism in a shallow Mediterranean macroalgal Meadow (*Caulerpa prolifera*). *Est. Coasts* 35, 1182–1192. doi: 10.1007/s12237-012-9514-y
- Velimirov, B. (1986). DOC dynamics in a Mediterranean seagrass system. *Mar. Ecol. Prog. Ser.* 28, 21–41. doi: 10.3354/meps028021
- Vonk, J. A., Middelburg, J. J., Stapel, J., and Bouma, T. J. (2008). Dissolved organic nitrogen uptake by seagrasses. *Limnol. Oceanogr.* 53, 542–548. doi: 10.4319/lo.2008.53.2.0542
- Wada, S., and Hama, T. (2013). The contribution of macroalgae to the coastal dissolved organic matter pool. *Est. Coast. Shelf Sci.* 129, 77–85. doi: 10.1016/j.ecss.2013.06.007
- Waycott, M., Duarte, C. M., Carruthers, T. J. B., Orth, R. J., Dennison, W. C., Olyarnik, S., et al. (2009). Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proc. Natl. Acad. Sci. U.S.A.* 106, 12377–12381. doi: 10.1073/pnas.0905620106
- Wetzel, R. G., and Penhale, P. A. (1979). Transport of carbon and excretion of dissolved organic carbon by leaves and roots/rhizomes in seagrasses and their epiphytes. *Aquat. Bot.* 6, 149–158. doi: 10.1016/0304-3770(79)90058-5
- Ziegler, S., and Benner, R. (1999). Dissolved organic carbon cycling in a subtropical seagrass-dominated lagoon. *Mar. Ecol. Prog. Ser.* 180, 149–160.
- Ziegler, S., Kaiser, E., and Benner, R. (2004). Dynamics of dissolved organic carbon, nitrogen and phosphorus in a seagrass meadow of Laguna Madre. *Texas. Bull. Mar. Sci.* 75, 391–407.

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